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# Analysis of Glycosylated Type II Interleukin-1 Receptor (IL-1R) by Imaged Capillary Isoelectric Focusing (i-cIEF)

Mei Han

Analytical Sciences

Amgen, Inc., Washington

CE Pharm 2007, Hyatt Regency, Miami

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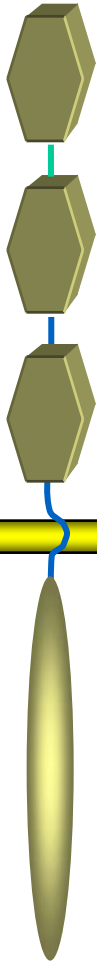
# Overview

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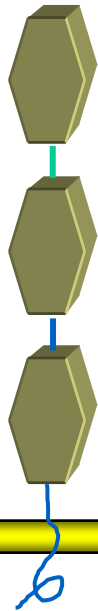
- **Introduction**
- **i-cIEF Method Development**
- **i-cIEF Applications**

# IL-1 Receptors

IL-1R  
Type I



IL-1R  
Type II

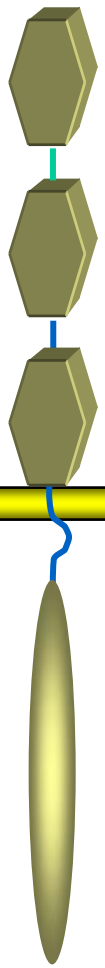


IL-1R  
AcP

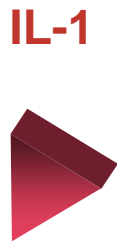


# IL-1 Binding

IL-1R  
Type I

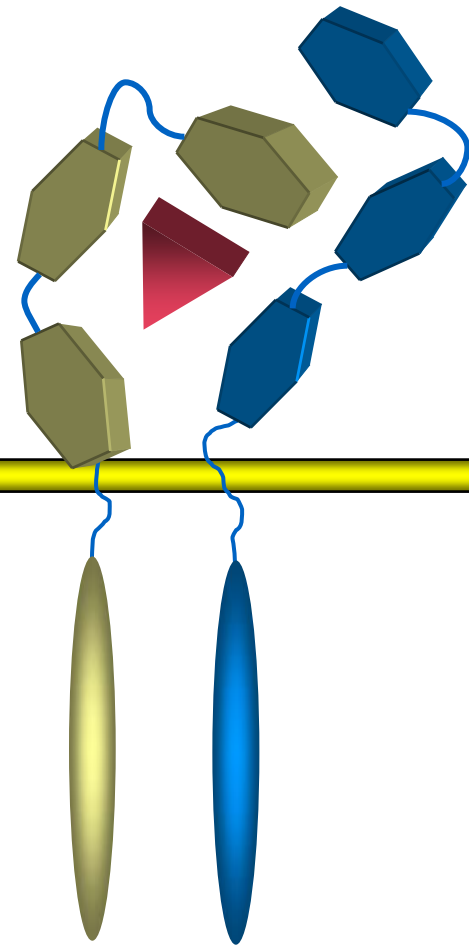
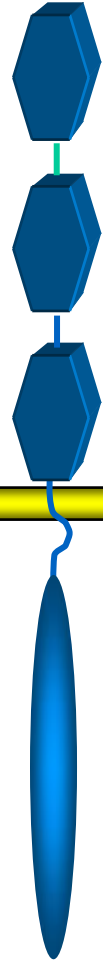


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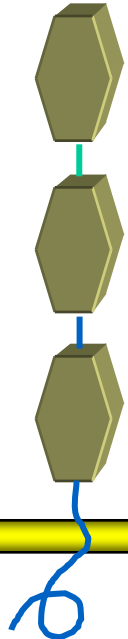
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IL-1R  
AcP



# IL-1 Receptor Type II is a Decoy Receptor

IL-1R  
Type II

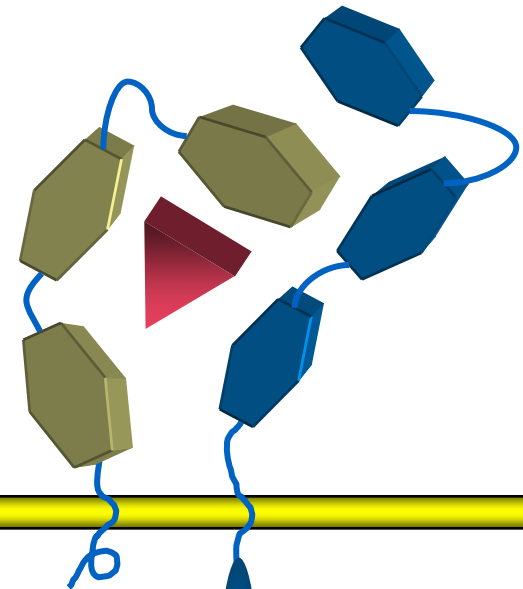


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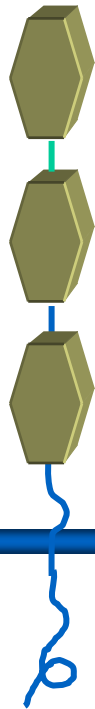
IL-1R  
AcP



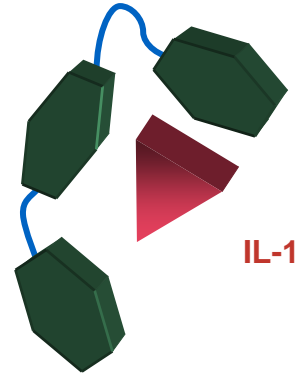
*Complex does not transfer signal:  
Type II IL-1R natural inhibitor of IL-1 biology*

# Soluble IL-1 Receptor Type II

IL-1R  
Type II



Soluble  
IL-1R Type II



*Main indication: rheumatoid arthritis (RA)*

# IL-1R Type II Molecule



- CHO expressed IL-1R II
- 333 Amino acids
- Peptide mass = 38,031 Da
- 4 disulfide bridges
- 5 Potential N-glycosylation sites
- Theoretical pI = 6.77
- Experimental pI range: 4.0-6.6
- Monomer in solution
- Glycoprotein mass = 46,520 Da

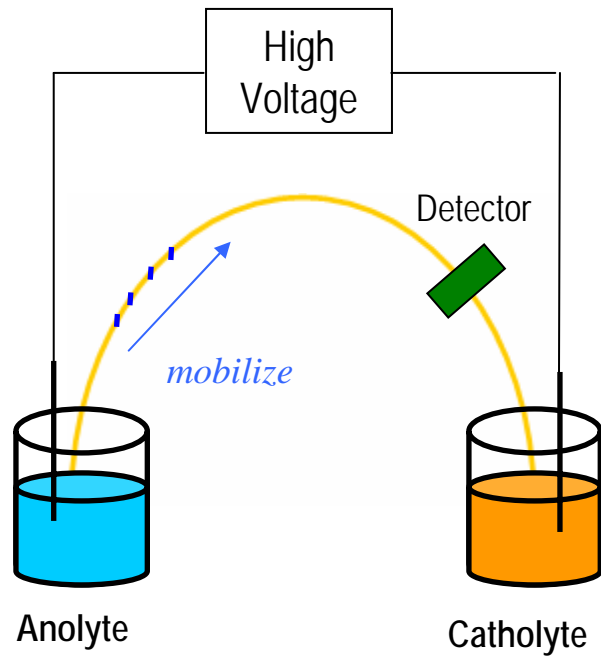
# Overview

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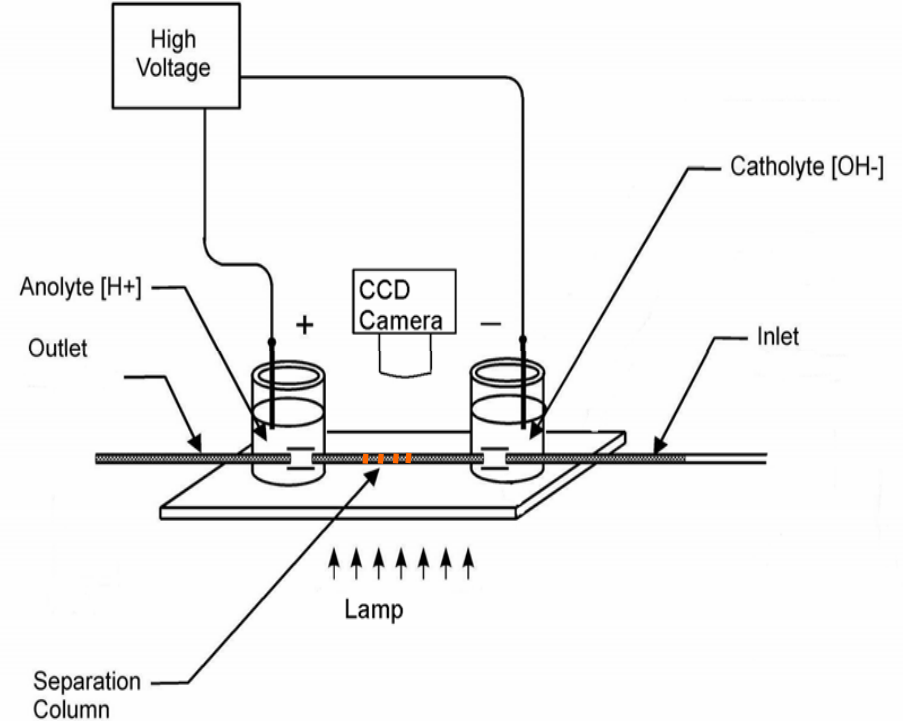
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# Imaged cIEF vs. Conventional cIEF

## Conventional cIEF



## Imaged cIEF (i-cIEF)



# Experimental

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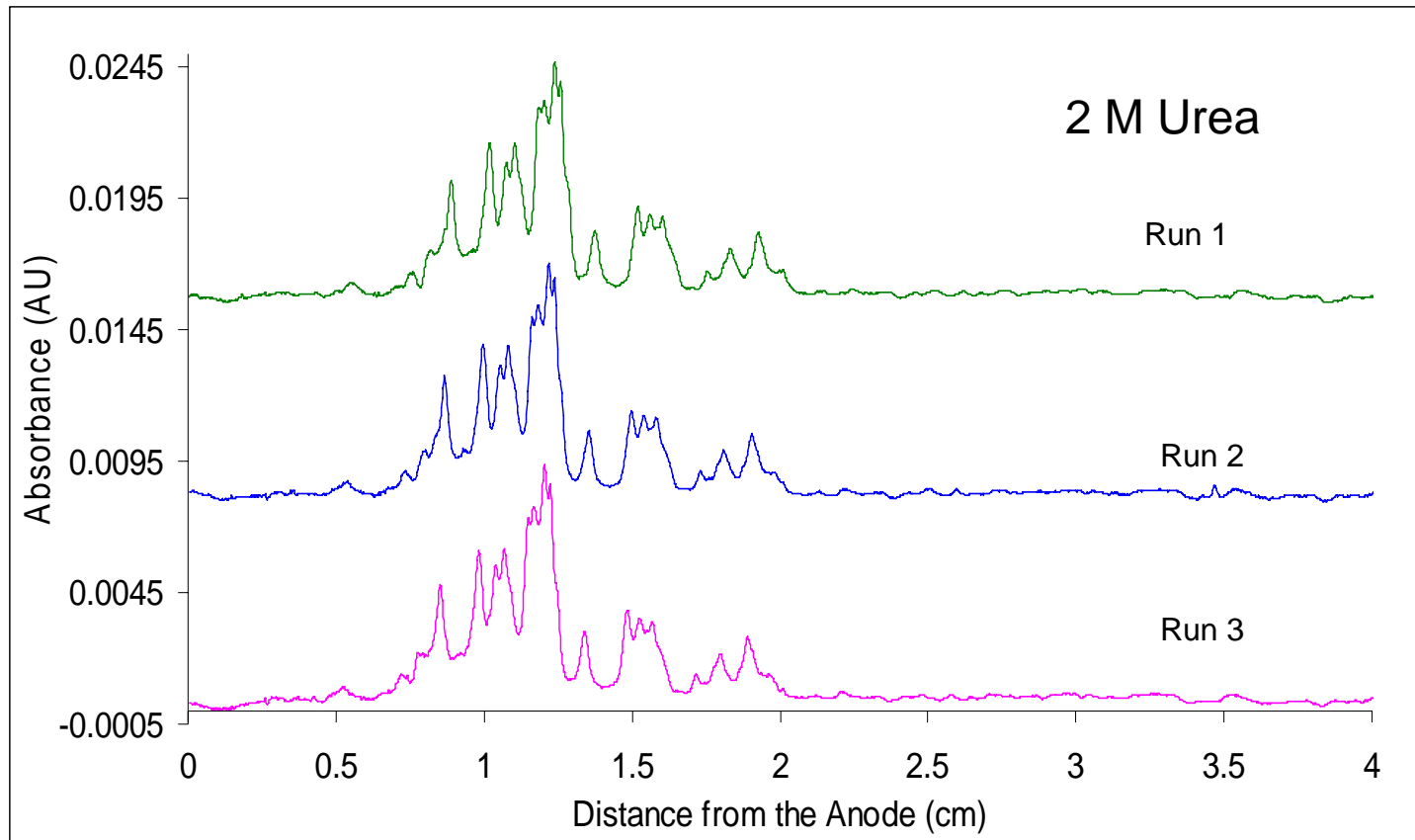
- **iCE280 w/Alcott autosampler Model 714 (Convergent Biosciences).**
- **Capillary cartridge : 5 cm x 100  $\mu$ m ID, FC coated (Convergent Biosciences).**
- **Detection: 280 nm.**
- **Software: iCE280 and EZChrom version 6.8**

# Method Development

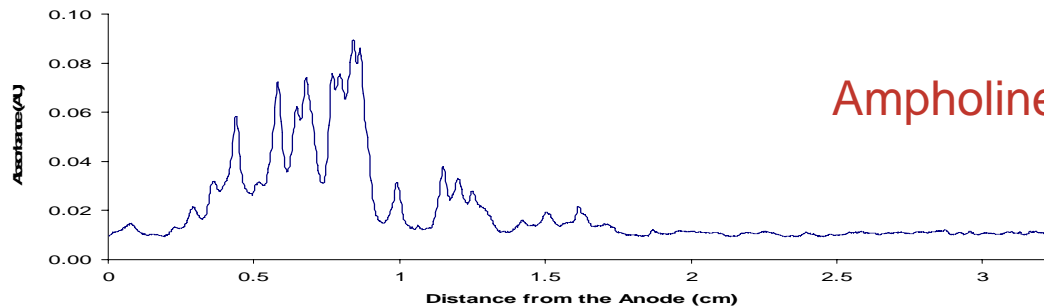
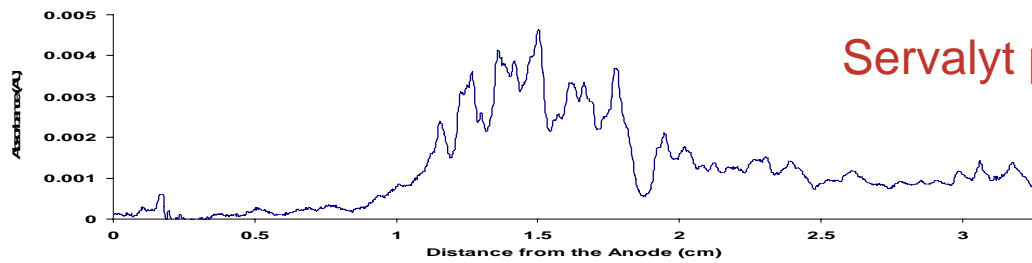
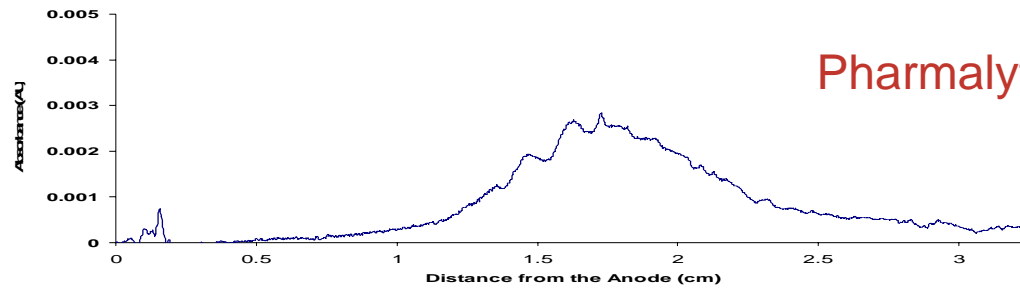
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- **Cartridge pre-condition: different concentrations (0.35% vs. 0.50% methyl cellulose) at different times (2, 4,... and 24 hours)**
- **Different anolyte and catholyte concentrations**
- **Different ampholytes: servalyt pH 2-11, pharmalyte 3-10, Ampholines pH 3.5-9.5 and mixed with narrow pH ranges**
- **Different additives and different urea final concentrations: 0, 1, 2, 3, 4 and 8 M**
- **Different focusing time: 4, 5, 6 and 7 minutes**
- **Injection linearity**
- **Intermediate precision**
- **Identity**

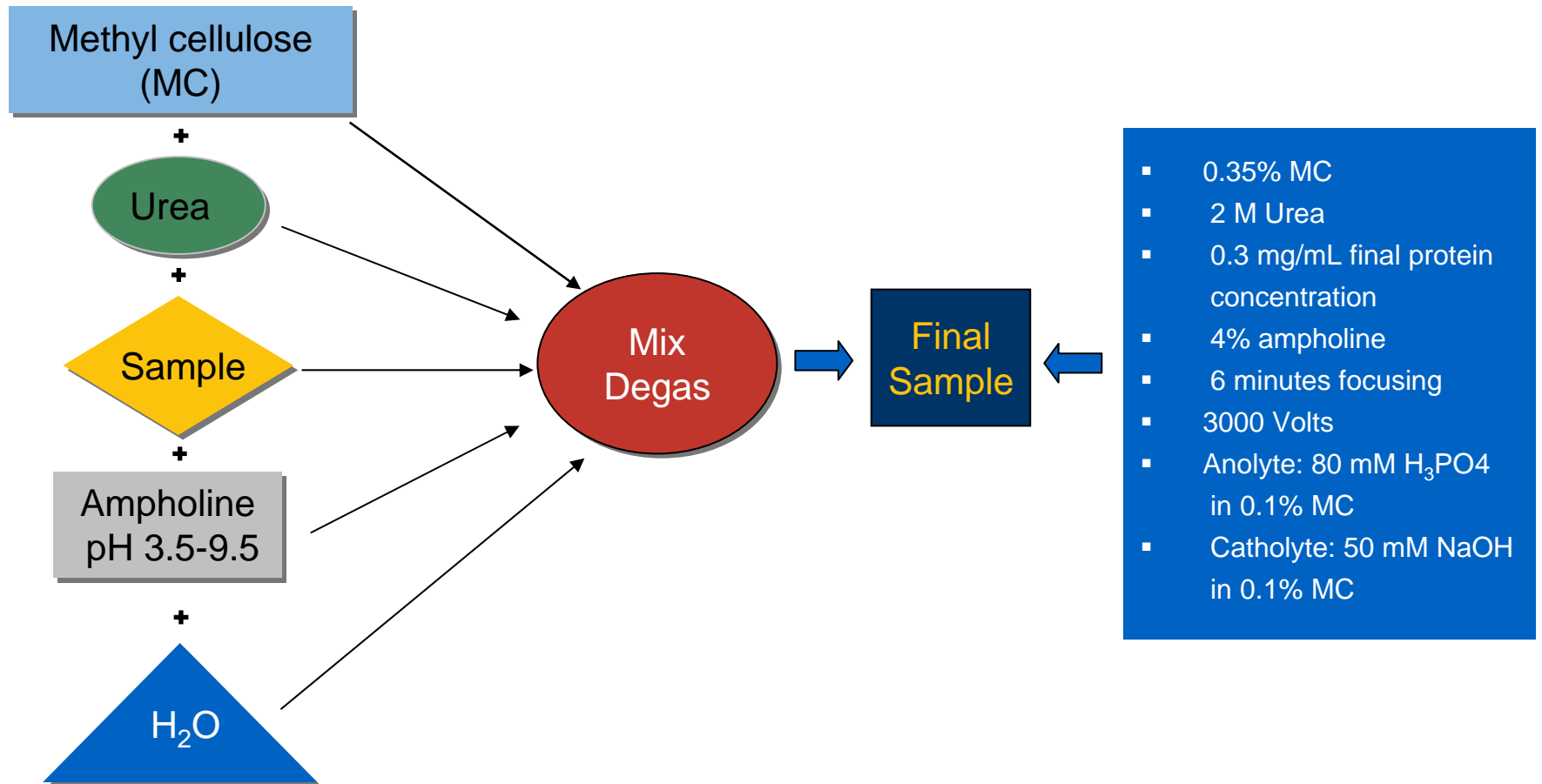
# Obtaining Repeatable Profiles



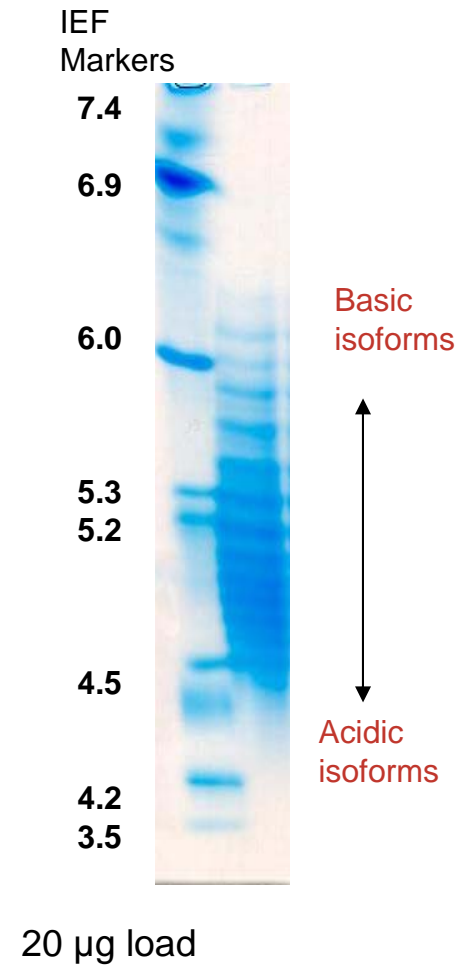
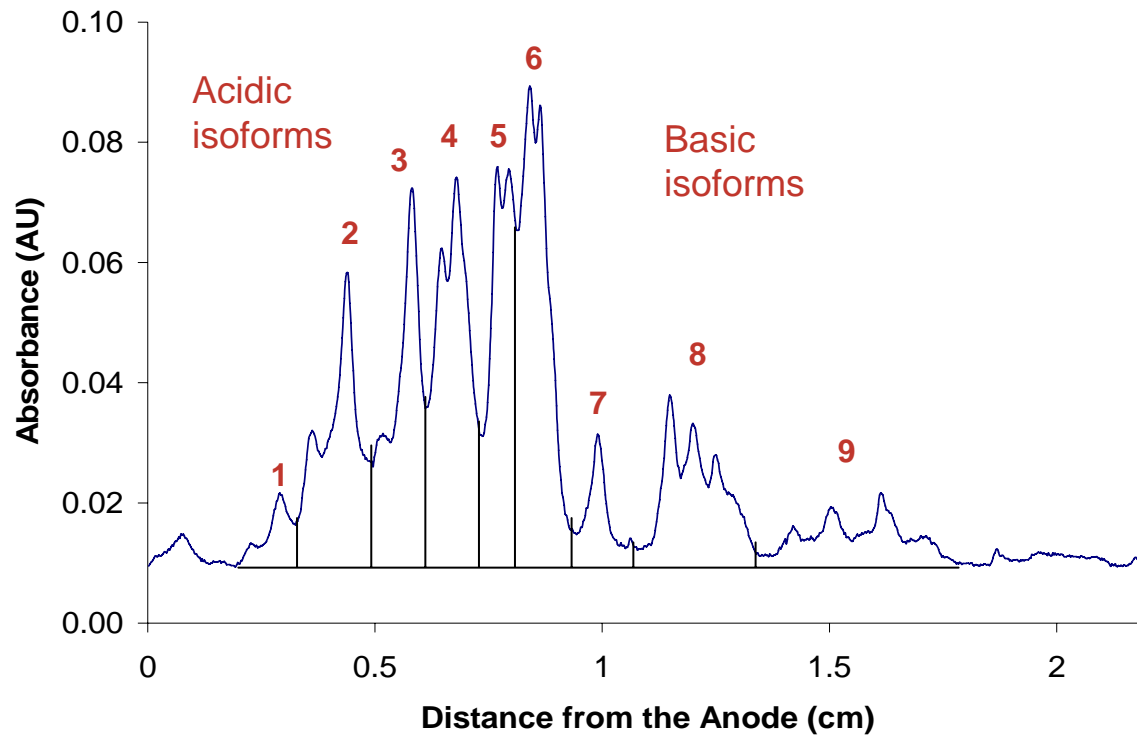
# Optimizing Resolution



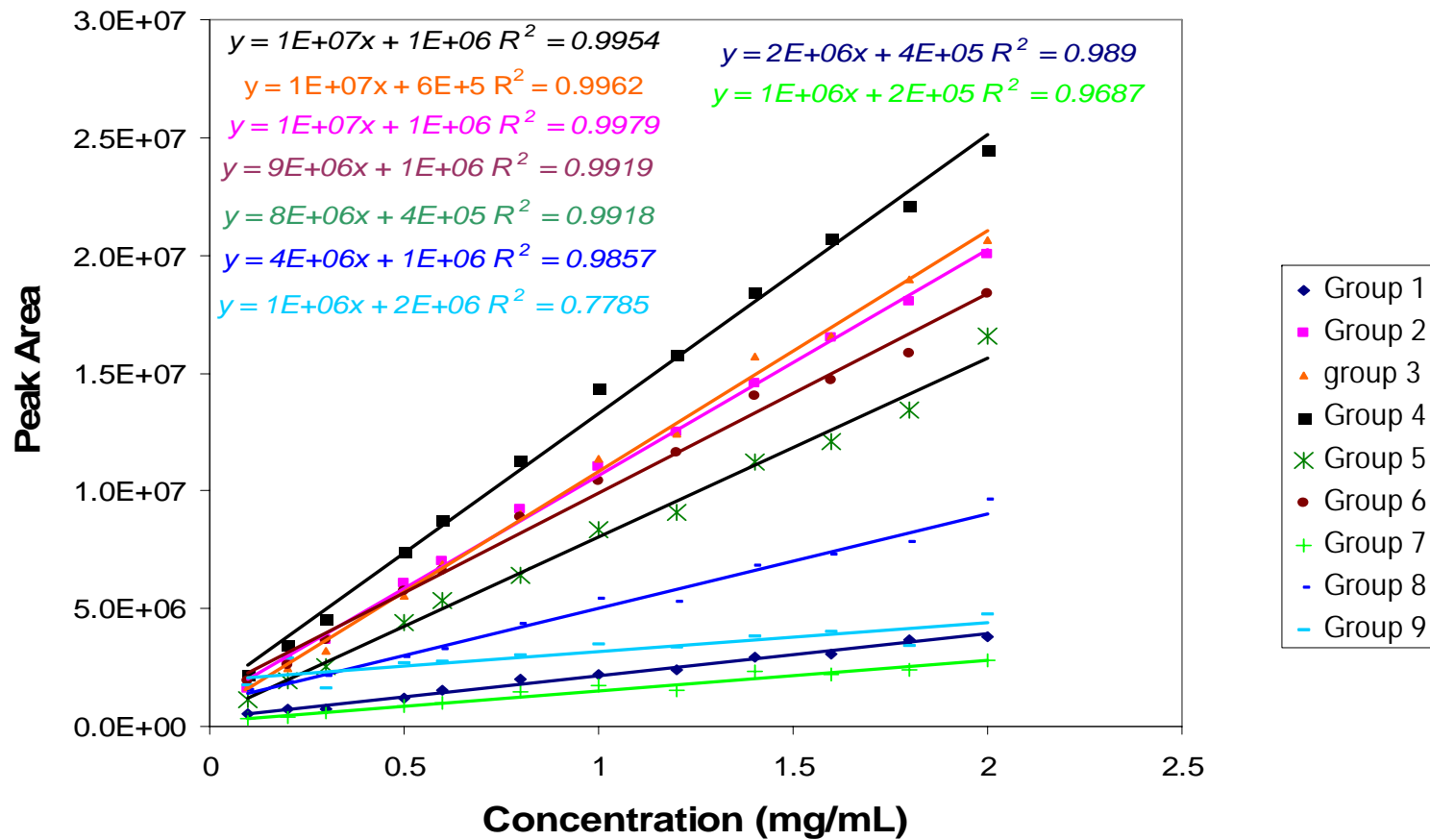
# Final Sample Preparation Condition



# Typical i-cIEF Profile Compared to the Slab Gel



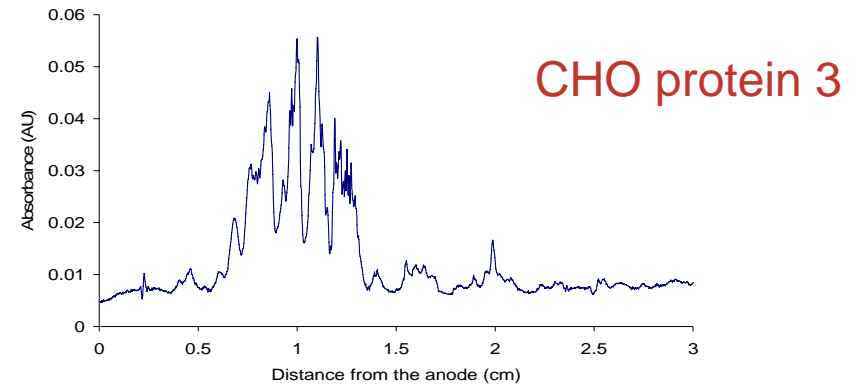
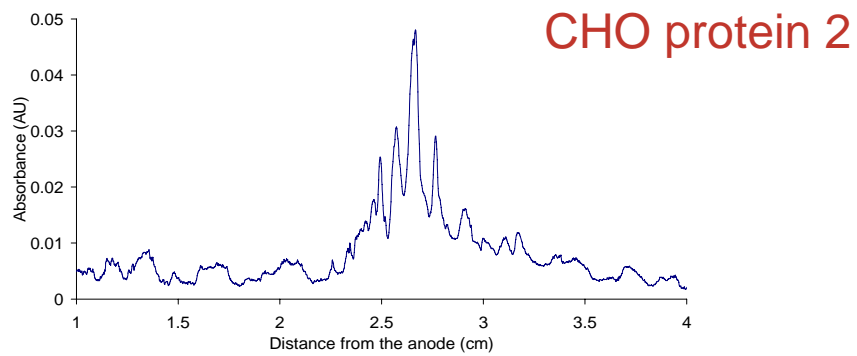
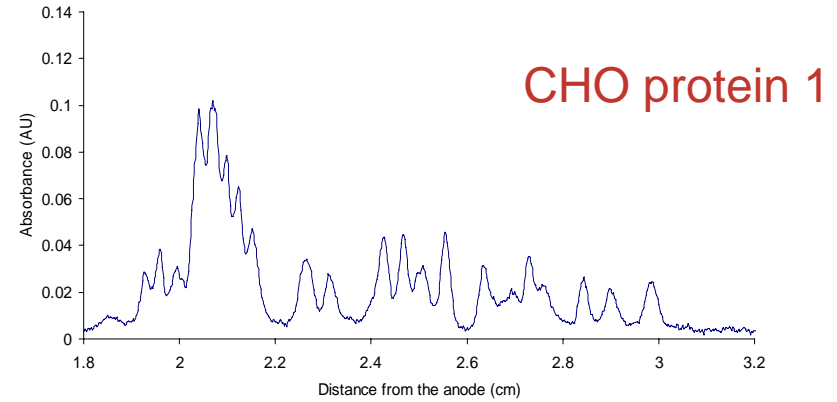
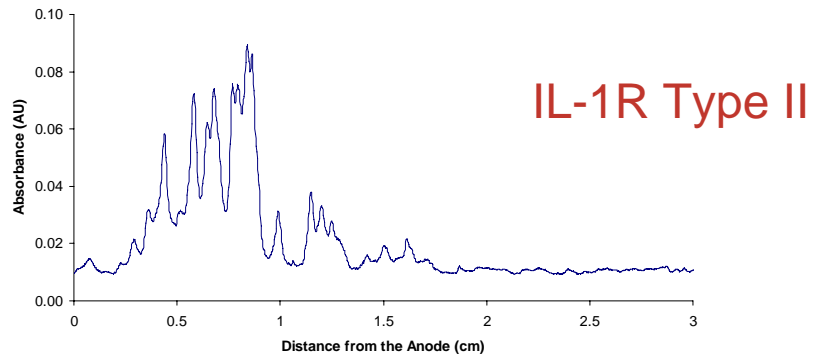
# Linearity Study of IL-1R II Charge Isoforms



# Intermediate Precision

Run #	% Peak Area								
	1	2	3	4	5	6	7	8	9
1	3.53	17.34	16.21	22.34	12.29	15.59	2.46	6.86	3.40
2	3.39	16.73	15.95	21.85	12.01	15.80	2.43	7.46	4.38
3	3.23	16.95	14.97	21.51	12.14	15.56	3.00	7.78	4.86
4	3.31	16.59	14.70	21.51	12.93	16.63	2.91	8.04	3.38
5	3.61	16.38	14.70	19.46	12.09	15.77	3.55	9.07	5.37
6	3.66	16.67	14.36	19.07	12.78	15.45	3.63	9.51	4.87
7	3.77	17.73	14.28	20.40	13.11	14.94	3.32	8.20	4.28
8	3.81	18.28	13.64	19.16	12.75	15.77	3.20	7.96	5.45
9	3.06	15.57	13.86	20.13	13.39	16.00	3.40	8.72	5.86
<b>Average</b>	3.49	16.92	14.74	20.60	12.61	15.72	3.10	8.18	4.65
<b>STD</b>	0.26	0.79	0.87	1.24	0.50	0.45	0.44	0.82	0.87
<b>%RSD</b>	7.32	4.68	5.89	6.00	3.93	2.88	14.14	9.99	18.78

# Identity

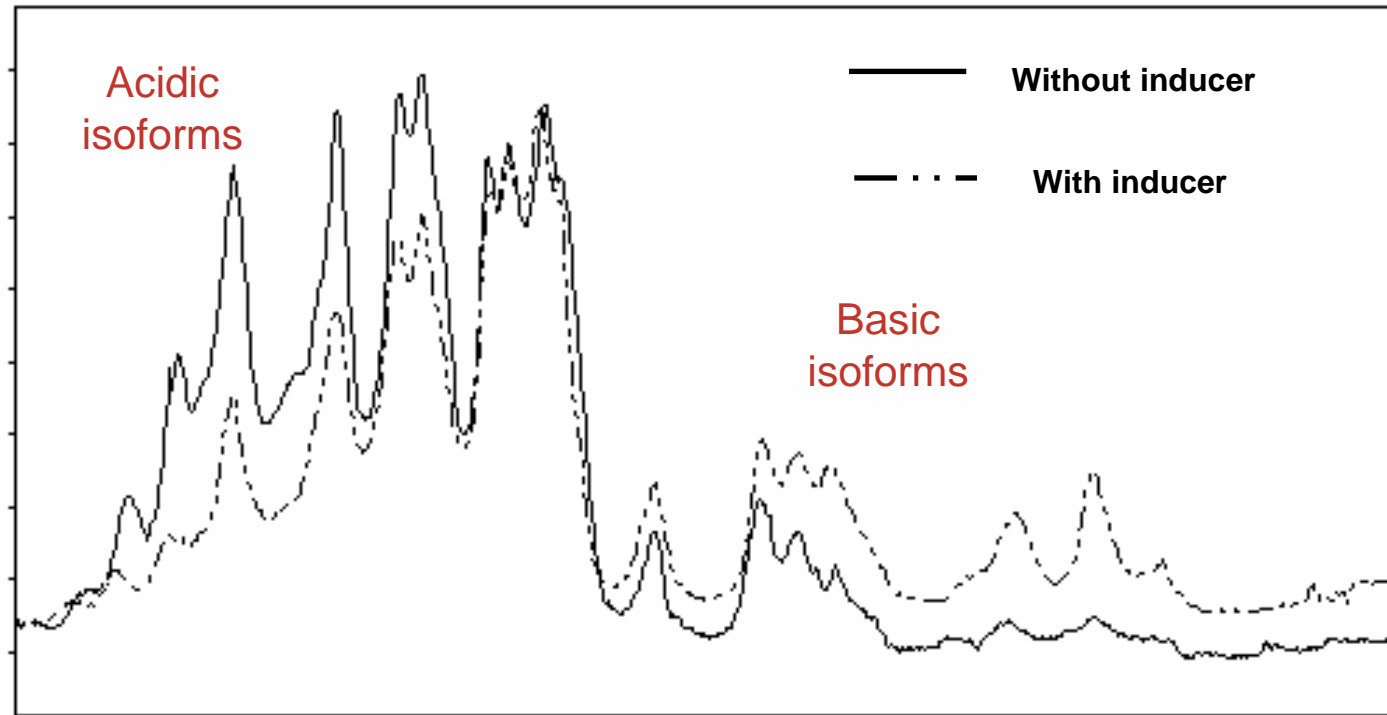


# Overview

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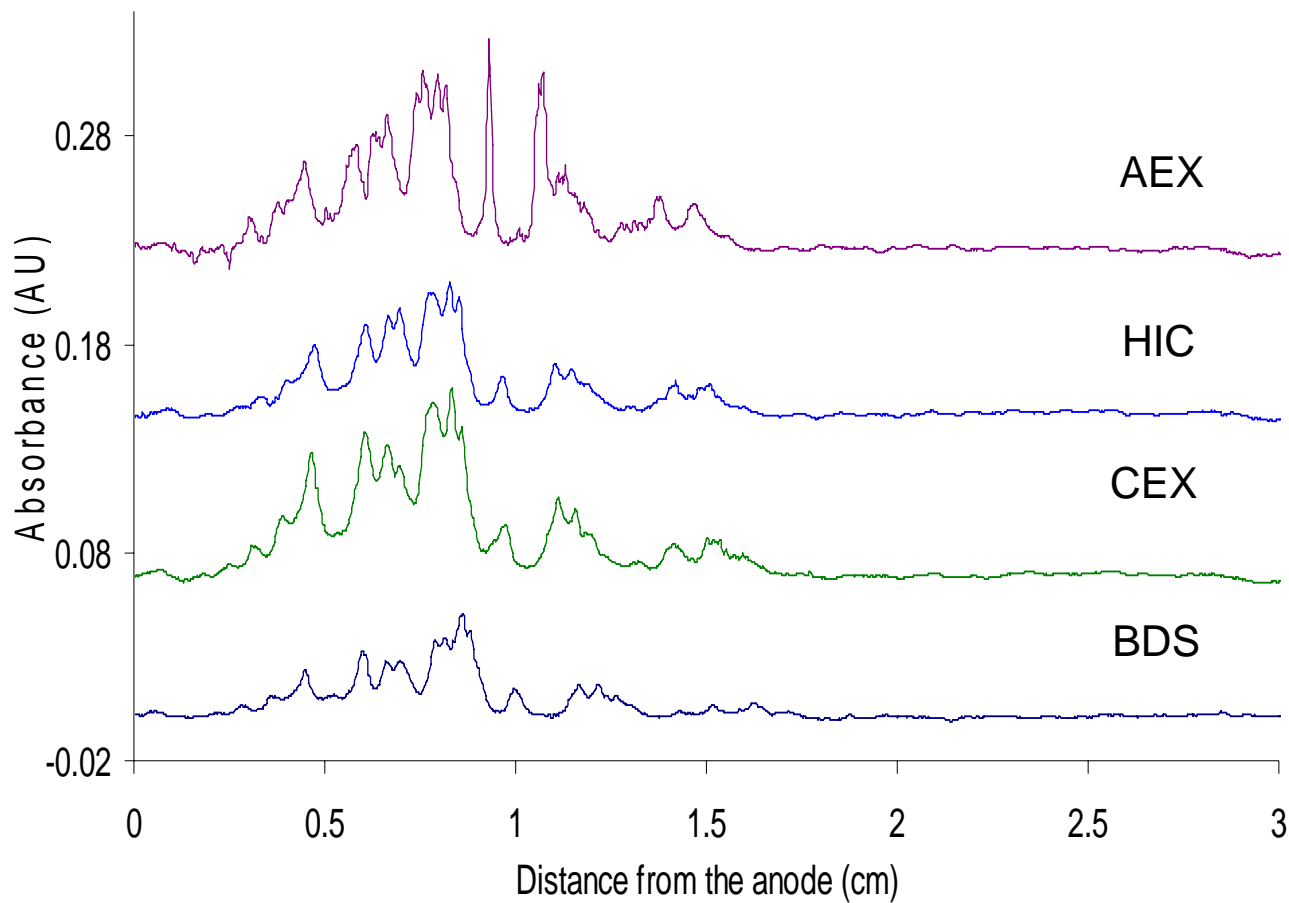
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# Effect of Inducer on i-cIEF Profile



	% Peak Area								
	1	2	3	4	5	6	7	8	9
w/o inducer	3.19	17.5	17.4	21.7	13.6	15.8	2.09	6.44	2.36
w/ inducer	1.59	9.40	11.9	19.3	15.0	16.6	3.79	12.9	9.44

# Monitoring Downstream Purification Process



Harvest

AEX



HIC

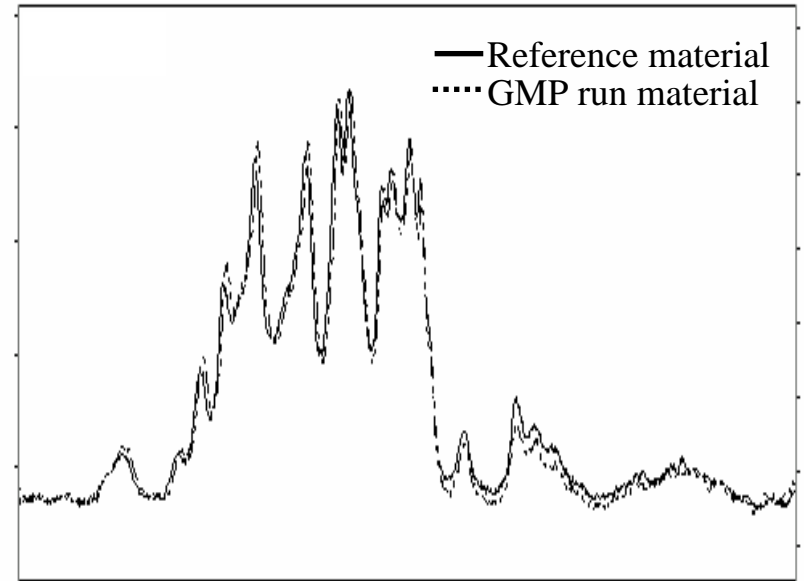
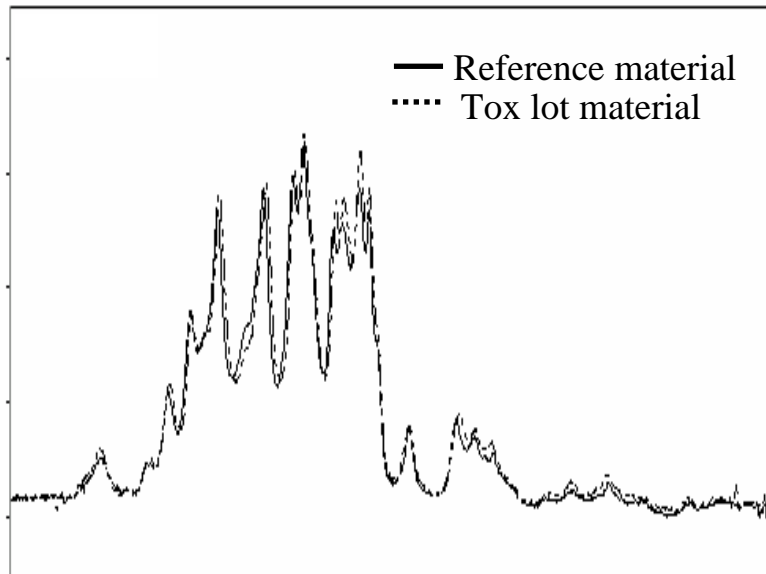


CEX

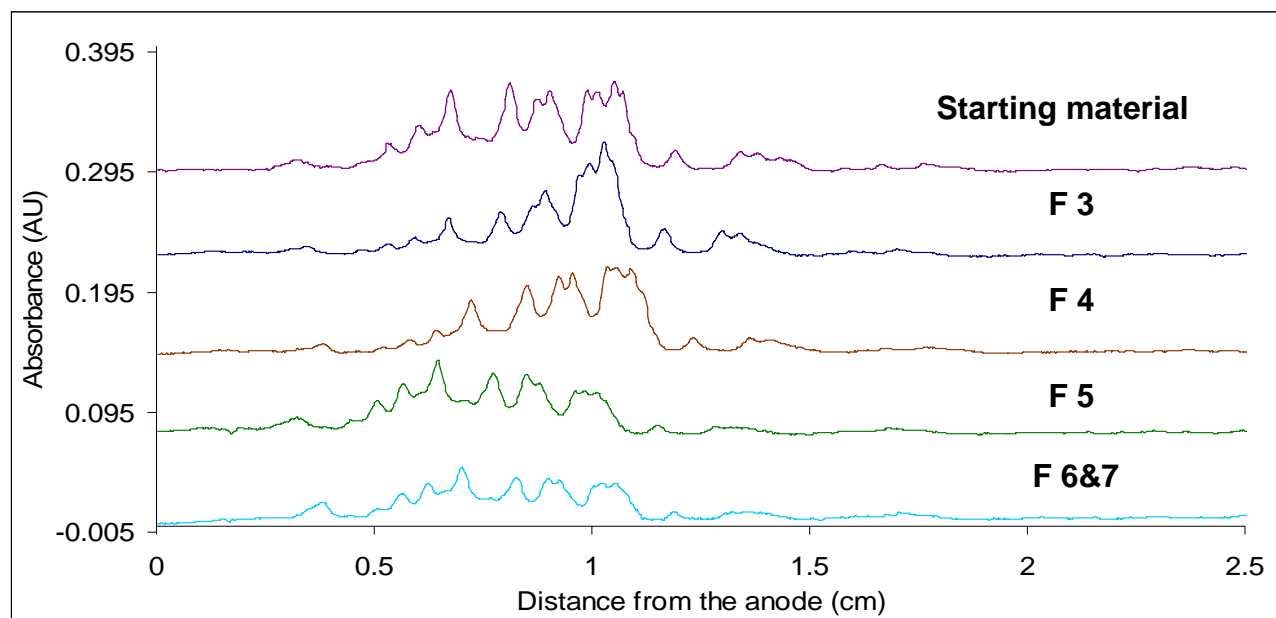
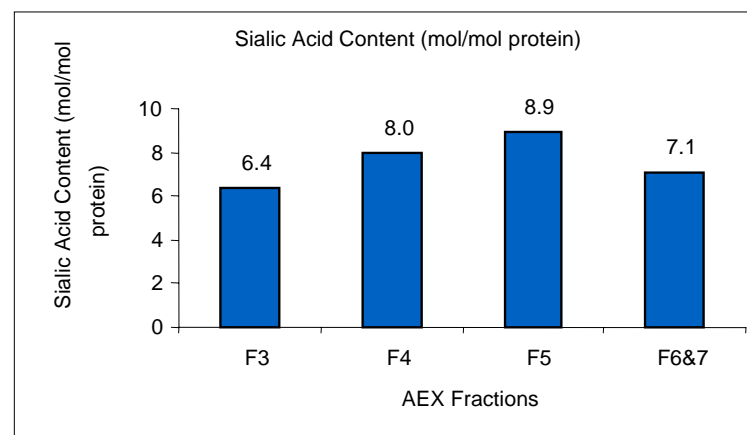
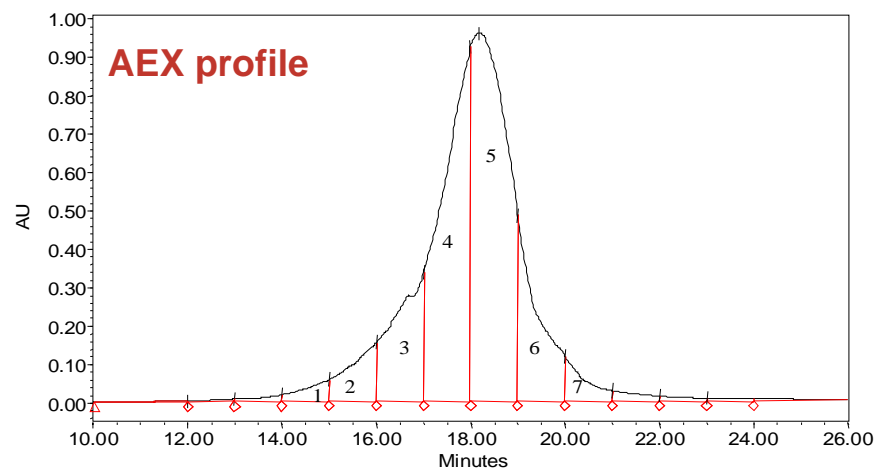


BDS

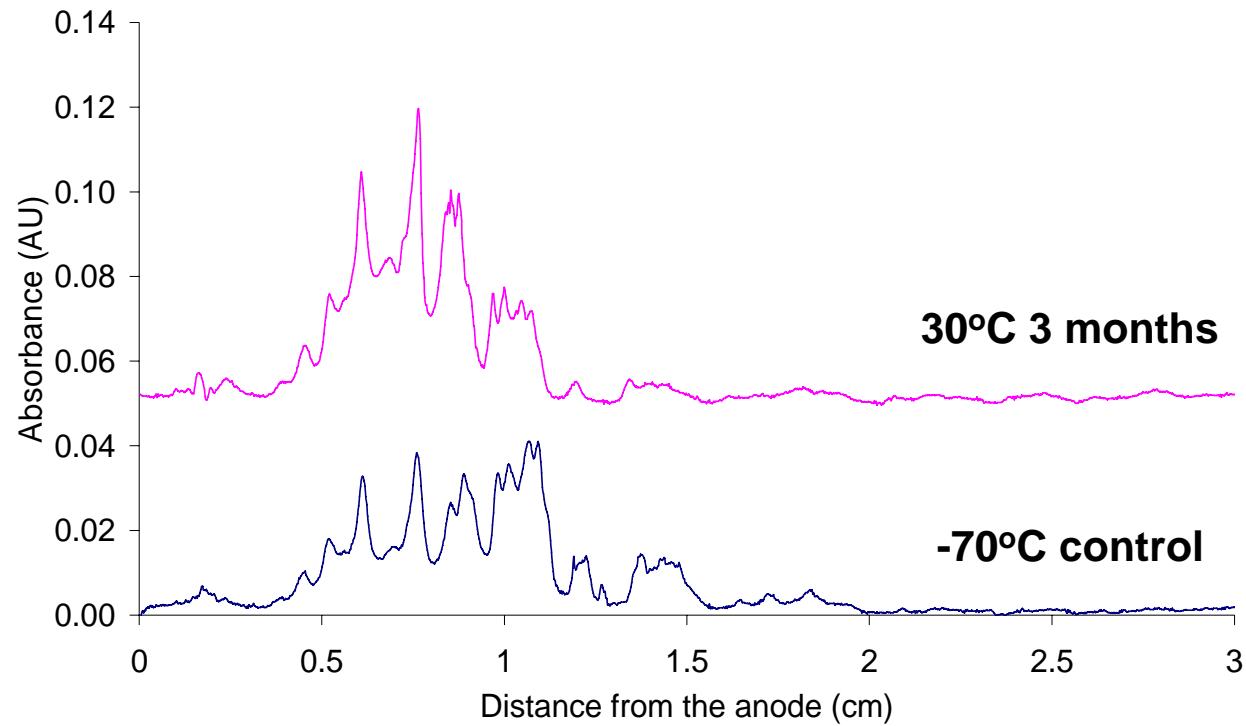
# Assessment of Lot-to-Lot Reproducibility



# Monitoring Sialic Acid Distribution



# i-cIEF As a Stability Indicating Assay



	% Peak Area								
	1	2	3	4	5	6	7	8	9
-70°C control	2.43	15.27	14.80	16.45	12.52	17.53	4.62	10.80	5.58
30°C 3 months	3.47	22.43	27.41	21.63	7.92	8.19	1.25	3.97	3.74

# Characterization: Stability Sample

Sample: Cyanogen Bromide (CNBr): 500:1



0.1 N HCl, 24 hours at RT

Dry out to remove unreacted CNBr and reduce molecule



25 mM DTT in presence of 3M  
GnHCl/0.1 M Tris pH 8.0

RP-HPLC/MS

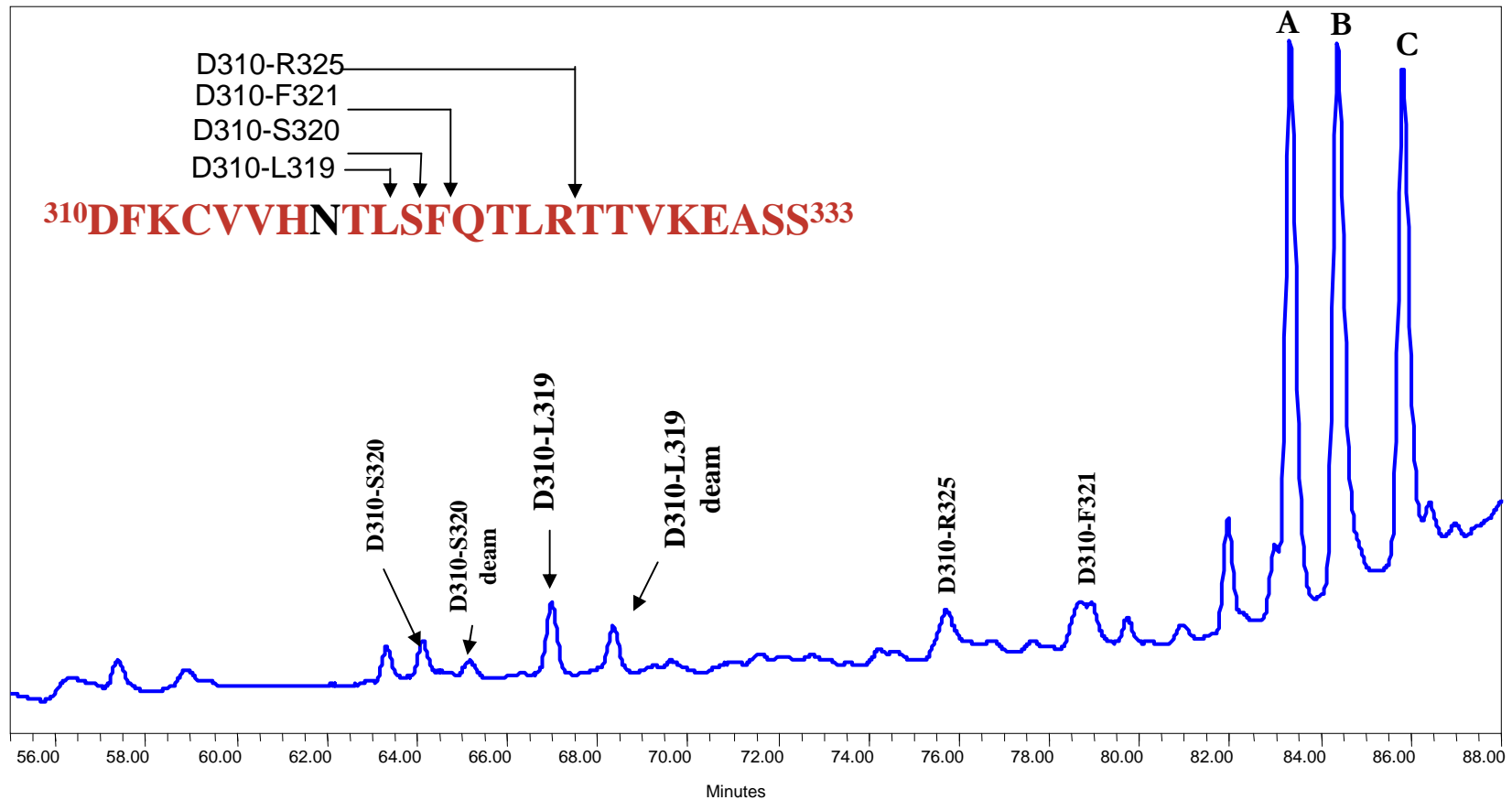
peaks collection



MALDI/TOF/MS analysis

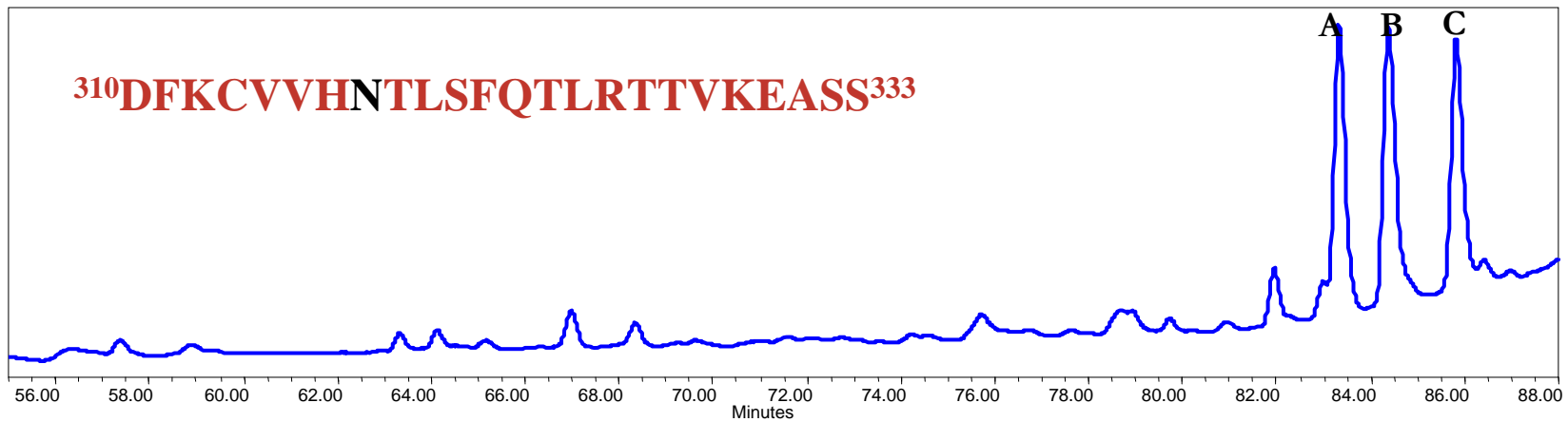
N-terminal Sequencing Analysis

# LC/MS Results of CNBr Digest of Stability Sample



30°C, 3 months sample

# MALDI/TOF/MS Results



CNBr Fractions	Theoretical Mass	Experimental Mass	Fragment (Modification)
A	2710.37	2711.28	D310-S333 (Isomerization)
B	2710.37	2710.28	D310-S333 (Unmodified)
C	2710.37	2711.27	D310-S333 (Deamidation)

# N-terminal Sequencing Results

CNBr RP-HPLC fractions	NTS Results	Modification
Fraction A	X-V-V-H (sequence stopped)	isomerization
Fraction B	X-V-V-H-N-T-L-S-F-Q-T-	unmodified
Fraction C	X-V-V-H-D-T-L-S-F-Q-T-	deamidation

# Conclusions

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- **i-cIEF analysis is a fast and automated method to monitor IL-1R Type II charge profile.**
- **This assay has a large linear response range and the profiles are reproducible. Organizing the isoforms in series of groups helps comparative and quantitative analyses.**
- **The distinctive electropherogram can be used as an identity fingerprint.**
- **i-cIEF is an effective method for monitoring upstream and downstream process.**
- **Bulk Drug Substance (BDS) as well as in process samples can be analyzed by this technique.**
- **i-cIEF is an appropriate method for monitoring deamidation.**

# Acknowledgements

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